

A new species of *Lasiobelba* from Nepal, with overview of the genus

(Acari, Oribatida, Oppiidae)

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A new species of the genus *Lasiobelba* (Oribatida, Oppiidae) – *L. sandormahunkai* sp. nov. – is described, based on materials collected from soil-litter in *Quercus semecarpifolia* forest of Nepal. Generic diagnosis and an identification key to all known species of *Lasiobelba* are presented. Taxonomic status of the related genera *Antennoppia*, *Lasiobelba* and *Oppia* and systematic placement of some representatives from them are discussed.

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Introduction

The oribatid mite genus *Lasiobelba* (Acari, Oribatida, Oppiidae) was proposed by Aoki (1959) with *Lasiobelba remota* Aoki, 1959 as type species. According to Subías (2004, online version 2021), the genus comprises two subgenera and 38 species, which have a cosmopolitan distribution collectively. The main goals of the paper are: to describe and illustrate a new species of *Lasiobelba* collected from Nepal; to summarize generic morphological traits; to discuss the taxonomic status of the genera *Lasiobelba*, *Antennoppia* Mahunka, 1983 and *Oppia* Koch, 1835 and systematic placement of some representatives from them, and to provide an identification key to all known species of *Lasiobelba*.

Presently, three species of *Lasiobelba* have been registered in Nepalese fauna (Ermilov & Martens 2014a,b, Ermilov et al. 2014): *L. daamsae* Ermilov, Shtanchaeva, Subías & Martens, 2014; *L. remota* Aoki, 1959; and *L. nepalica* Ermilov, Shtanchaeva, Subías & Martens, 2014.

Material and methods

Specimens. Oribatid mites were collected from the Jochen Martens Expeditions to Nepal in 1988¹ and until recently were housed in the Institut für Organismische und Molekulare Evolutionsbiologie (Mainz, Germany).

Observation and documentation. Specimens were mounted in lactic acid on temporary cavity slides

¹ Results of the Himalaya Expeditions of J. Martens from 1969–2004, No. 293. Jochen Martens was sponsored by DAAD and DFG.

for measurement and illustration. Body length was measured in dorsal view, from the tip of the rostrum to the posterior edge of the notogaster. Notogastral width refers to the maximum width of the notogaster in dorsal view. Lengths of body setae were measured in lateral aspect. All body measurements are presented in micrometers. Formulas for leg setation are given in parentheses according to the sequence trochanter-femur-genu-tibia-tarsus (familus included). Formulas for leg solenidia are given in square brackets according to the sequence genu-tibia-tarsus. Drawings were made with a camera lucida using a Leica transmission light microscope "Leica DM 2500".

Terminology. Morphological terminology used in this paper follows that of F. Grandjean: see Travé & Vachon (1975) for references, Norton (1977) for leg setal nomenclature, and Norton & Behan-Pelletier (2009), for overview.

Abbreviations. Prodorsum: *ro, le, in, bs, ex* = rostral, lamellar, interlamellar, bothridial, and exobothridial seta, respectively; *mt* = median tubercle; *pt* = postbothridial tubercle. Notogaster: *c* = setal alveolus; *la, lm, lp, h, p* = setae; *ia, im, ip, ih, ips* = lyrifissures; *gla* = opisthontal gland opening. Gnathosoma: *a, m, h* = subcapitular setae; *or* = adoral seta; *d, l, v, cm, ul, su, vt, lt* = palp setae; *w* = palp solenidion; *cha, chb* = cheliceral setae; *Tg* = Trägårdh's organ. Epimeral and lateral podosomal regions: *1a-1c, 2a, 3a-3c, 4a-4c* = epimeral setae; *PdI* = pedotectum I; *dis* = discidium. Anogenital region: *g, ag, an, ad* = genital, aggenital, anal, and adanal seta, respectively; *iad* = adanal lyrifissure; *po* = preanal organ. Legs: *Tr, Fe, Ge, Ti, Ta* = trochanter, femur, genu, tibia, and tarsus, respectively; *ω, φ, σ* = solenidia; *ε* = famulus; *d, l, v, bv, ev, ft, tc, it, p, u, a, s, pv, pl* = setae; *pa* = porose area.

Taxonomy

Family Oppiidae

Subfamily Oppiinae

Genus *Lasiobelba* Aoki, 1959

Type species: *Lasiobelba remota* Aoki, 1959

Main generic traits of *Lasiobelba* (adult)

Size. Length about 230–1300.

Integument. Mostly smooth; lateral side usually densely tuberculate; sometimes prodorsum and/or notogaster heavily wrinkled or granulate/tuberculate.

Prodorsum. Rostrum rounded/pointed/tripartite/truncate. Costula and transcostula absent; lateral prodorsal carina present or absent. Interbothridial and postbothridial tubercles present or absent. Interbothridial region with or without muscle sigillae.

Fig. 1. *Lasiobelba sandormahunkai* sp. nov., adult: A. dorsal view (not shown: legs); B. ventral view (not shown: ▷ gnathosoma and legs); C. right lateral view (not shown: gnathosoma and legs); D. posterior view. Scale bar 200 µm.

Rostral, lamellar, interlamellar, and exobothridial setae usually well developed, setiform; rarely, some of them very short or represented by alveolus; *le* inserted closer to *in* or to *ro*. Bothridial seta spindle-form, with long setiform or elongate conical apex.

Notogaster. Without humeral tooth and crista. Usually ten pairs of setiform setae; *c* short or represented by alveolus; rarely, seta *c* and/or *h₃* absent (eight or nine pairs of notogastral setae present in these cases).

Gnathosoma. Subcapitulum diarthric. Adoral seta present, setiform. Palp setation: 0-2-1-3-8 or 9(+1 solenidion). Palp solenidion short, in medio-anterior part close to distal seta. Chelicera chelate-dentate.

Epimeral and lateral podosomal regions. Epimeral border IV present. Epimeral setal formula: 3-1-3-3; all setae setiform. Ventrosejugal tubercle absent. Pedotectum I represented by small lamina. Discidium present.

Anogenital region. Five pairs of genital, one pair of aggenital, two pairs of anal, and three pairs of adanal setae; all setae setiform. Adanal seta *ad₁* posterior or posterolateral, *ad₂* lateral, *ad₃* anterolateral to anal plate; distance between *ad₃-ad₃* larger than *ag-ag*, and *ad₂-ad₂*. Adanal lyrifissure close and parallel to anal aperture.

Legs. Tarsus I with 20 setae (*l''* and *v'* present), tarsus II with 15 or 16 setae (*l''* present or absent). Tarsus II with two solenidia.

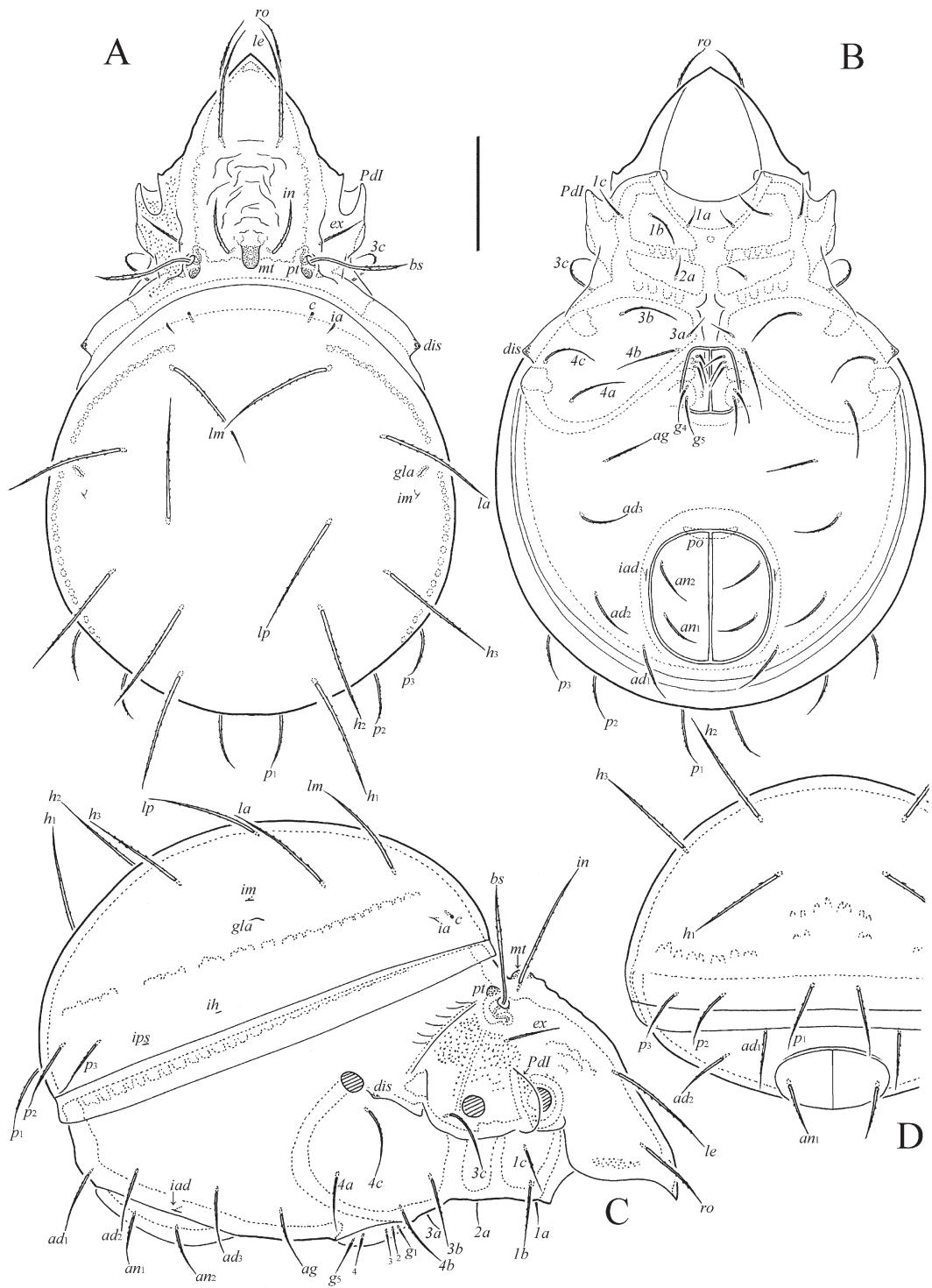
Lasiobelba sandormahunkai sp. nov.

Figs 1-3

Diagnosis. Body size: 1095–1162 × 713–747. Prodorsum dorsally wrinkled. Rostrum pointed. Rostral, lamellar, interlamellar, and exobothridial setae long, setiform, barbed; *in*>*le*>*ro*>*ex*. Bothridial seta long, with slightly developed head, barbed. Interbothridial muscle sigillae absent. Interbothridial region with one strong unpair tubercle. Nine pairs of long, setiform, barbed notogastral setae (*p₁-p₃* shorter than others); tenth pair of setae (*c*) represented by alveoli. Rutellum with mediodial tooth. Epimeral and anogenital setae setiform, mostly slightly barbed. Discidium present. Adanal lyrifissure longitudinal.

Description

Measurements. Very large species. Body length: 1095 (holotype: male), 1095–1162 (13 paratypes, sex



not identified); notogaster width: 713 (holotype), 713–747 (13 paratypes).

Integument. Body colour brown. Body surface mostly smooth. Prodorsal region between lamellar and interlamellar setae wrinkled. Lateral part of body between bothridium and acetabula I–III densely tuberculate (diameter of tubercle up to 4).

Prodorsum. Rostrum pointed, but often broken, therefore it may appear truncated or rounded. Rostral (131–139), lamellar (184–205), interlamellar (215–233), and exobothridial (94–98) setae setiform, barbed; *ex* thinnest. Bothridial seta (200–215) with slightly developed, elongate, lanceolate head, barbed; conical apex distinct. Interbothridial muscle sigillae not observed. Interbothridial region with one strong unpair tubercle located between interlamellar setae. Postbothridial tubercle present. Longitudinal row, comprising several muscle sigillae, present in front of the bothridium.

Notogaster. Anterior border convex medially. Nine pairs of notogastral setae (p_1-p_5 : 110–114; others: 192–205) setiform, barbed; tenth pair of setae (c) represented by alveoli. All notogastral lyrifissures (except *ip* not observed) and opisthonotal gland opening distinct.

Gnathosoma. Subcapitulum size: 225–246 × 184–200. Rutellum with mediol distal tooth. Subcapitular setae (a: 49–57; m: 82–94; h: 94–106) setiform, barbed. Adoral seta (28–32) setiform, smooth. Palp length: 159–168. Solenidion short, bacilliform, located in dorsodistal part of tarsus. Postpalpal seta (10) spiniform, smooth. Chelicera length: 225–246. Cheliceral setae (cha: 69–73; chb: 49–53) setiform, barbed.

Epimeral and lateral podosomal regions. All epimeral setae (1a, 2a, 3a: 32–41; 1b: 94–102; 1c: 73–77; 3b, 3c, 4a, 4b, 4c: 110–123) setiform, slightly barbed. Discidium strong, triangular.

Anogenital region. Genital setae (g_1-g_3 : 32–41; g_4 : 45–53; g_5 : 61–69) setiform, roughened. Aggenital, adanal and anal setae (110–114) setiform, barbed. Adanal lyrifissure distinct.

Legs. All leg claws slightly barbed on dorsal side. Porose area on femora I–IV slightly visible. Formulas of leg setation and solenidia: I(1-5-2-4-

20) [1-2-2], II(1-5-2-4-16) [1-1-2], III(2-3-1-3-15) [1-1-0], IV(1-2-2-3-12) [0-1-0]; homology of setae and solenidia indicated in Table 1. Famulus of tarsus I erect, slightly swollen and blunted distally, inserted between solenidion ω_1 and seta ft'' . Seta p eupathidial on tarsus I, and conical on tarsi II–IV. Seta s eupathidial on tarsus I, located before setae a' and a'' . Solenidion ω_1 on tarsus I, ω_1 and ω_2 on tarsus II slightly thickened, rounded apically, others setiform.

Material examined. Holotype and 13 paratypes: Nepal, Katmandu Distr., Sheopuri Mt., 27°47'N, 85°23'E, 2100–2300 m a.s.l., soil-litter in *Quercus semecarpifolia* forest, 25.VI.1988 (collected by J. Martens and W. Schwaller).

Type deposition. The holotype and two paratypes are deposited in the collection of the Senckenberg Museum of Natural History, Görlitz, Germany; 11 paratypes are deposited in the collection of the Tyumen State University Museum of Zoology, Tyumen, Russia. All specimens are preserved in ethanol with a drop of glycerol.

Etymology. The new species is named after late Prof. Dr. S. Mahunka (1937–2012), the Hungarian acarologist, for his extensive contributions to our knowledge of fauna and taxonomy of oribatid mites.

Remarks. The new species differs from the all known species of the genus by the presence (versus absence or presence of one pair) of one unpair tubercle between interlamellar setae.

Discussion

The oribatid mite genus *Oppia* was proposed by Koch (1835) with *Oppia nitens* Koch, 1835 as type species. Later, Aoki (1959) proposed the genus *Lasiobelba* with *Lasiobelba remota* Aoki, 1959 as type species, and Mahunka (1983) proposed the genus *Antennoppia* with *Antennoppia minor* Mahunka, 1983 as type species. These genera differ from each other by one main generic trait (bothridial seta clavate/fusiform/lanceolate, without long apex in *Oppia*;

Table 1. Leg setation and solenidia of *Lasiobelba sandormahunkai* sp. nov. Roman letters refer to normal setae, Greek letters to solenidia (except ε=famulus); single quotation mark (') designates setae on the anterior and double quotation mark (") setae on the posterior side of a given leg segment; parentheses refer to a pair of setae.

Leg	Tr	Fe	Ge	Ti	Ta
I	v'	d, (l), bv'', v''	(l), σ	(l), (v), ϕ ₁ , ϕ ₂	(ft), (tc), (it), (p), (u), (a), s, (pv), (pl), l'', v', ε, ω ₁ , ω ₂
II	v'	d, (l), bv'', v''	(l), σ	(l), (v), φ	(ft), (tc), (it), (p), (u), (a), s, (pv), l'', ω ₁ , ω ₂
III	l', v'	d, l', ev'	l', σ	l', (v), φ	(ft), (tc), (it), (p), (u), (a), s, (pv)
IV	v'	d, ev'	d, l'	l', (v), φ	ft'', (tc), (p), (u), (a), s, (pv)

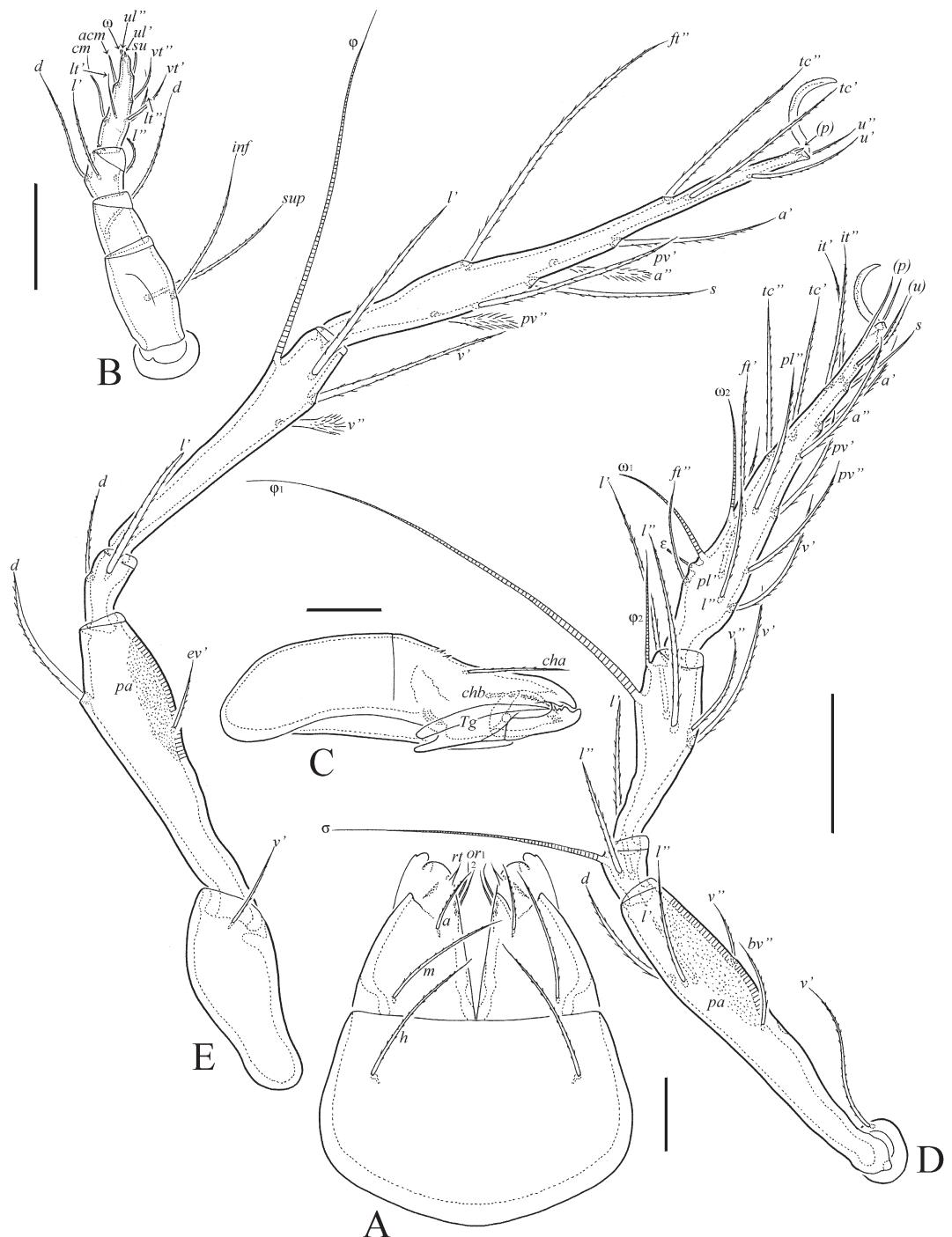


Fig. 2. *Lasiobelba sandormahunkai* sp. nov., adult: A. subcapitulum, ventral view; B. palp, left, paraxial view; C. chelicera, left, paraxial view; D. leg I, right, antiaxial view; E. leg IV, left, antiaxial view. Scale bar 50 µm (A,C), 100 µm (D,E).

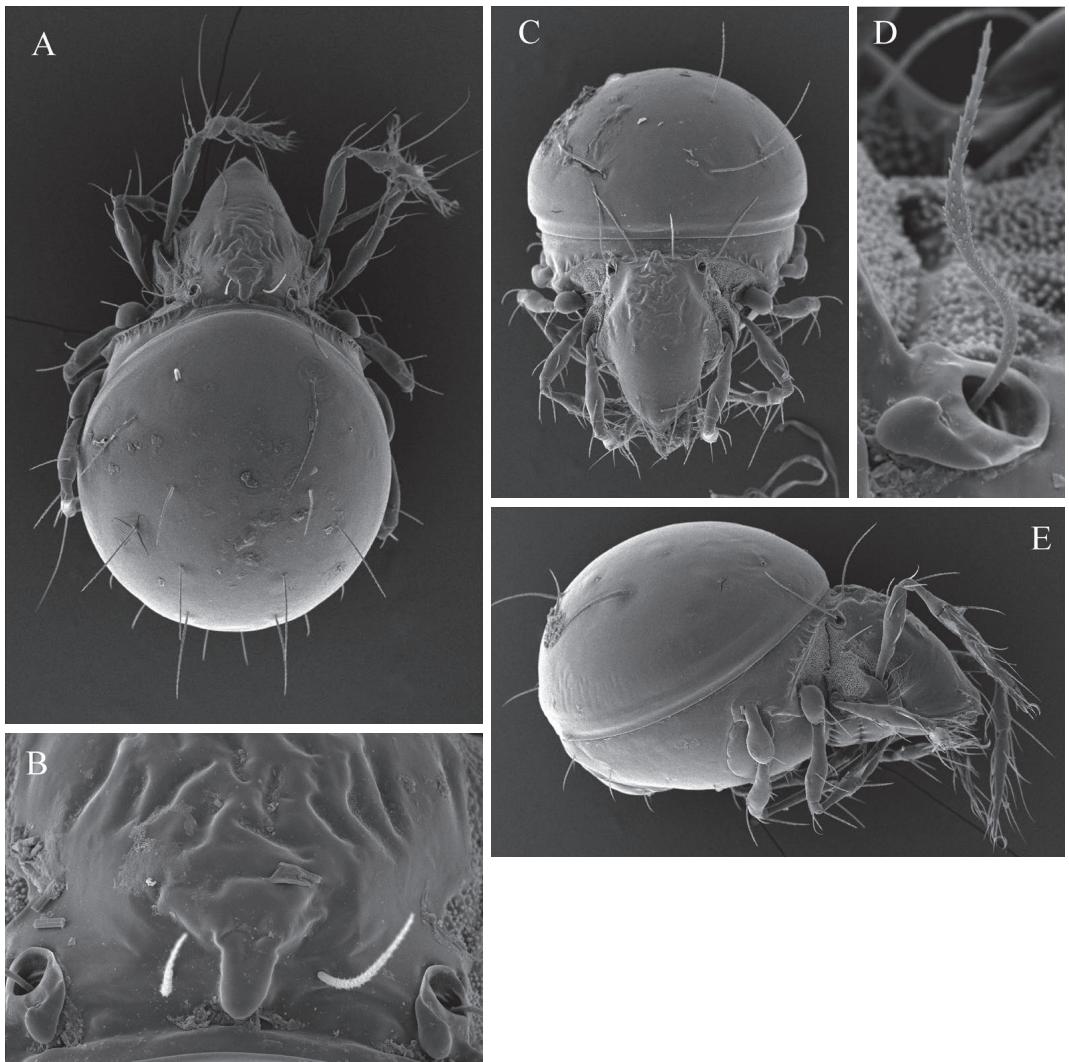


Fig. 3. *Lasiobelba sandormahunkai* sp. nov., adult, SEM micrographs: **A.** dorsal view; **B.** mediobasal part of prodorsum, dorsal view; **C.** dorsoanterior view; **D.** bothridial seta, right, lateral view; **E.** lateral view.

spindle-form having long conical/setiform apex in *Lasiobelba*; setiform in *Antennoppia*), however, in general appearance they are morphologically very similar, therefore, the opinion on their taxonomic status was different. For example, Subías & Balogh (1989) synonymized *Antennoppia* with *Lasiobelba*; Subías (2004) included *Antennoppia* as subgenus in *Lasiobelba*; Balogh & Balogh (2002) included representatives of *Lasiobelba* and *Antennoppia* in *Oppia*, implying the synonymy of them.

So far, in relation to the above listed genera, the Subías's system (2004) is supported, in which *Oppia* and *Lasiobelba* are independent genera, and *Anten-*

noppia is the subgenus of *Lasiobelba* (e.g. Ermilov et al. 2014, Ermilov & Friedrich 2016, Subías online version 2021). However, in our current opinion, if these genera differ only in morphology of bothridial seta, then it is unjustifiable to support the different taxonomic status for *Oppia*, *Lasiobelba* and *Antennoppia* (genus versus subgenus); it would be more logical for all them to support the initial generic statuses or, as an alternative, to include *Lasiobelba* and *Antennoppia* as subgenera in *Oppia* with the formation of subgenera *Oppia* (*Oppia*), *Oppia* (*Lasiobelba*) and *Oppia* (*Antennoppia*).

Thus, we decided to preliminarily support (and reject our early opinions – e.g. Ermilov et al. 2014, Ermilov & Friedrich 2016) the taxonomic independence of genera *Oppia*, *Lasiobelba* and *Antennoppia* until additional studies (e.g. molecular or phylogenetic) are presented.

After an explanation above listed, we were able to identify the incorrect taxonomic placement of several species of *Lasiobelba*, because morphology of their bothridial seta corresponds to the generic diagnosis of *Oppia*, therefore, we offer combinations for these species: *O. decui* (Vasiliu & Ivan, 1995) comb. nov., *O. hesperidiana* (Pérez-Íñigo, 1986) comb. nov., *O. pontica* (Vasiliu & Ivan, 2011) comb. nov., *O. sakhalinensis* (Ryabinin & Zaitsev, 2019) comb. nov., and *O. suchetae* (Sanyal, 1992) comb. nov. Initial generic placement for *Oppia arcidiaconoae* Bernini, 1973 and *O. kuehnelti* Csiszár, 1961 (= *O. yodai* Aoki, 1965, = *O. yodai africana* Kok, 1967, = *Cilioppia pori* Vasiliu & Ivan, 1995, = *Lasiobelba arabica* Mahunka, 2000, = *L. neonominata* Subías, 2004)² is supported.

Also, we support initial systematic placement for some members of *Antennoppia* (*A. granulata* Mahunka, 1986, *A. major* Mahunka, 1983, *A. minor* Mahunka, 1983, *A. trichoseta* Mahunka, 1983, and *A. yoshii* Mahunka, 1987) and offer combinations for other *Antennoppia*-species from *Lasiobelba* (*Antennoppia*) in *Antennoppia*: *A. abchasica* (Golosova & Tarba, 1974) comb. nov., *A. capilligera* (Berlese, 1916) comb. nov., *A. chistyakovi* (Ermilov & Kalúz, 2012) comb. nov., *A. heterosa* (Wallwork, 1964) comb. nov., *A. insignis* (Balogh, 1970) comb. nov., *A. izquierdoae* (Arillo, Gil-Martín & Subías, 1994) comb. nov., *A. nepalica* (Ermilov, Shtanchaeva, Subías & Martens, 2014) comb. nov., *A. parachistyakovi* (Ermilov, 2016) comb. nov. (in Ermilov & Friedrich 2016), *A. quadrisetata* (Subías, 1989) comb. nov. (in Subías & Balogh 1989), *A. rigida* (Ewing, 1909) comb. nov., *A. subnitida* (Sellnick, 1924) comb. nov., and *A. ultraciliata* (Jacot, 1934) comb. nov.

Key to known species of *Lasiobelba*

After the taxonomic acts proposed in the previous section, *Lasiobelba* comprises 14 species, which have distributed collectively in the Ethiopian and Oriental region, Argentina, Japan and Santa Helena Island.

1. Notogastral setae *la* and *lm*³ distinctly shorter than bothridial seta..... 2
- Notogastral setae *la* and *lm* longer than bothridial seta, or they slightly differ in length.... 7
2. Dorsal notogastral setae comparatively short (*la* and *lm* distinctly not reaching insertion of *lp*) body length: 787–825 *Lasiobelba rubida* (Wallwork, 1977). Distribution: Santa Helena Island.
- Dorsal notogastral setae comparatively long (*la* or/and *lm* reaching insertion of *lp*)..... 3
3. Notogastral seta *c* short; body length: 478–522 *Lasiobelba lemuria* Mahunka, 1997. Distribution: Madagascar.
- Notogastral seta *c* represented by alveolus or absent 4
4. Surface of notogaster and dorsal part of prodorsum not wrinkled or granulate; lamellar seta shorter than rostral and interlamellar setae..... 5
- Surface of notogaster and dorsal part of prodorsum wrinkled or granulate; lamellar seta not shorter than rostral and interlamellar setae..... 6
5. Notogastral seta *h*₃ present; notogastral seta *h*₂ of medium length; body length: 237–246..... *Lasiobelba longisensilla* Ermilov, 2017. Distribution: Vietnam.
- Notogastral seta *h*₃ absent; notogastral seta *h*₂ short; body length: 265–315..... *Lasiobelba tsaoshanensis* Ermilov, 2018 (see Ermilov & Liao 2018). Distribution: Taiwan.

² Subías (2004, online version 2021) considered *Lasiobelba arabica* Mahunka, 2000 (= *Oppia yodai africana* Kok, 1967, = *Lasiobelba neonominata* Subías, 2004) and *Oppia kuehnelti* Csiszár, 1961 (= *Oppia yodai* Aoki, 1965, = *Cilioppia pori* Vasiliu & Ivan, 1995) as independent species.

³ There is confusion with the designation of notogastral setae *la* and *lm*. In our current opinion, if setae *la* and *lm* are not arranged in one longitudinal row, then *lm* is located anteromedially (often very clearly) or medially to *la*. Hence, designations of *la* and *lm* are not correct in some papers and must be swapped (e.g. Ermilov et al. 2014, Ermilov & Starý 2018).

6. Surface of notogaster and dorsal part of prodorsum wrinkled; interbothridial region with one pair of strong tubercles; body length: 693.....
.....*Lasiobelba sculptra*
Wang, 1993. Distribution: southeastern China.
- Surface of notogaster and dorsal part of prodorsum granulate; interbothridial region without one pair of strong tubercles; body length: 610-644.....
.....*Lasiobelba yunnanensis*
Wen, 1999. Distribution: southeastern China.
7. Rostrum tripartite; notogastral seta h_3 absent; body length: 464-514.....
.....*Lasiobelba camerunica*
Ermilov & Starý 2018. Distribution: Cameroon.
- Rostrum not tripartite; notogastral seta h_3 present 8
8. Rostrum pointed 9
- Rostrum not pointed 11
9. Surface of dorsal part of prodorsum wrinkled; interbothridial region with one unpair strong tubercle; body length: 481-531.....
.....*Lasiobelba sandormahunkai* sp. nov.
Distribution: Nepal.
- Surface of dorsal part of prodorsum not wrinkled; interbothridial region without one unpair strong tubercle 10
10. Anterior part of pedotectum I elongate, forming specifically curved tooth; notogastral setae p_1-p_3 longer than adanal setae; body length: 1278-1310.....
.....*Lasiobelba daamiae*
Ermilov, Shtanchaeva, Subías & Martens, 2014.
Distribution: Nepal.
- Anterior part of pedotectum I not elongate and not forming specifically curved tooth; notogastral setae p_1-p_3 not longer than adanal setae; body length: 772-891
.....*Lasiobelba gibbosa*
(Mahunka, 1985). Distribution: Ethiopian region.
11. Rostrum truncate with one pair slight lateral teeth; body length: 794-834
.....*Lasiobelba insulata*
Ohkubo, 2001. Distribution: Japan, Taiwan.
- Rostrum rounded 12
12. Interlamellar seta distinctly shorter than bothridial seta body length: 625-684
.....*Lasiobelba vietnamica*
Balogh, 1983 (as *Oppia remota* in Balogh & Mahunka 1967). Distribution: Vietnam.
- Interlamellar seta longer than bothridial seta, or they slightly differs in length 13
13. Notogastral setae p_1-p_3 located comparatively close to each other (distance p_3-p_3 equally to ad_2-ad_2 ; body length: 560
.....*Lasiobelba subuligera*
(Berlese, 1916) (see also Mahunka & Mahunka-Papp 1995). Distribution: Argentina.
- Notogastral setae p_1-p_3 clearly distanced from each other (distance p_3-p_3 larger than ad_2-ad_2 ; body length: 858-1030
.....*Lasiobelba remota*
Aoki, 1959 (see also Ohkubo 2001). Distribution: Oriental region and East of eastern Palaearctic region.

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References

- Aoki, J. 1959. Die Moosmilben (Oribatei) aus SüdJapan. Bulletin of the Biogeographical Society of Japan 21(1): 1-22.
- 1965. Oribatiden (Acarina) Thailands. I. Nature and Life in Southeast Asia 4: 129-193.
- Arillo, A., Gil-Martin, J. & Subías, L. S. 1994. Oribatidos del "M.S.S." de las Islas Canarias. Poroscheloribatinae subfam. n. (Acari, Oribatida). Mémoires de Biospéologie 21: 1-6.
- Balogh, J. 1970. New oribatids (Acari) from New Guinea. II. Acta Zoologica Academiae Scientiarum Hungaricae 16(3-4): 291-344.
- 1983. A partial revision of the Oppiidae Grandjean, 1954 (Acari: Oribatei). Acta Zoologica Academiae Scientiarum Hungaricae 29(1-3): 1-79.
- & Balogh, P. 2002. Identification keys to the oribatid mites of the Extra-Holarctic regions. Vol. 1. 453 pp., Miskolc (Well-Press Publishing Limited).
- & Mahunka, S. 1967. New oribatids (Acari) from Vietnam. Acta Zoologica Academiae Scientiarum Hungaricae 13(1-2): 39-74.

- Berlese, A. 1916. Centuria terza di Acari nuovi. Redia 12: 289–338.
- Bernini, F. 1973. Notulae oribatologicae VII. Gli Oribatei (Acarida) dell'isolotto di Basiluzzo (Isole Eolie). Lavori della Societa Italiana di Biogeografia, Nuova Serie 3: 355–480.
- Csiszár, M. J. 1961. New oribatids from Indonesian soils (Acari). Acta Zoologica Academiae Scientiarum Hungaricæ 7(3-4): 345–366.
- Ermilov, S. G. 2017. A new species of *Lasiobelba* (Acari, Oribatida, Oppiidae) from Vietnam. Ecologica Montenegrina 15: 33–39.
- & Friedrich, S. 2016. Additions to the oppioid oribatid mite fauna of Peru (Acari, Oribatida, Oppioidea). Acarologia 56(3): 379–391.
- & Kalúz, S. 2012. Two new species of Oppiidae (Acari: Oribatida) from Ecuador. International Journal of Acarology 38(6): 521–527.
- & Liao, J.-R. 2018. A new species of *Lasiobelba* (Acari, Oribatida, Oppiidae) from Taiwan. Acarina 26(1): 81–87.
- & Martens, J. 2014a. Additions to the Nepalese oribatid mite fauna, with description of two new species (Acari, Oribatida). International Journal of Acarology 40(2): 123–132.
- & Martens, J. 2014b. New species, new records and a checklist of oribatid mites (Acari: Oribatida) from Nepal. Biologia 69(12): 1716–1729.
- & Starý, J. 2018. Oribatid mites (Acari, Oribatida) from Korup National Park (Cameroon): list of taxa, new findings, descriptions of two new species. Systematic and Applied Acarology 23(4): 733–747.
- , Shtanchaeva, U. Ya., Subías, L. S. & Martens, J. 2014. Two new species of oribatid mites of *Lasiobelba* (Acari, Oribatida, Oppiidae) from Nepal, including a key to all species of the genus. ZooKeys 424: 1–17.
- Ewing, H. E. 1909. New American Oribatoidea. Journal of the New York Entomological Society 17(3): 116–136.
- Golosova, L. D. & Tarba, Z. M. 1974. New species and genera of the superfamily Oppioidea (Acariformes, Oribatei) from Abkhazia and Primorsky Krai. Zoolichesky Zhurnal 53(12): 1885–1887.
- Jacot, A. P. 1934. Some Hawaiian Oribatoidea (Acarina). Bernice P. Bishop Museum Bulletin, Honolulu 121: 1–99.
- Koch, C. L. 1835. Deutschlands Crustaceen, Myriapoden und Arachniden, Vol. 1–3. Regensburg.
- Kok, D. J. 1967. Studies on some South African Oppiidae Grandjean, 1953 (Acarina: Oribatei). Journal of the Entomological Society of Southern Africa 30(1): 40–74.
- Mahunka, S. 1983. Oribatids from the Eastern Part of the Ethiopian Region II. Acta Zoologica Academiae Scientiarum Hungaricæ 29(1–3): 151–180.
- 1985. Description and redescription of Ethiopian oribatids (Acari, Oribatida), II. Annales Historico-Naturales Musei Nationalis Hungarici 77: 233–249.
- 1986. Oribatids from Africa (Acari: Oribatida) III. Folia Entomologica Hungarica 47(1–2): 41–76.
- 1987. Neue und interessante Milben aus dem Genfer Museum LV. Oribatids from Sabah (East Malaysia) I (Acari: Oribatida). Archives des Sciences 40(3): 293–305.
- 1997. Oribatids from Madagascar III (Acari: Oribatida). (Acarologica Genavensis LXXXIII). Revue Suisse de Zoologie 104(1): 115–170.
- 2000. Some oribatid mites from Yemen (Acari: Oribatida) (Acarologica Genavensis LXXXVIII). Annales Historico-Naturales Musei Nationalis Hungarici 92: 325–346.
- & Mahunka-Papp, L. 1995. The oribatid species described by Berlese (Acari). 325 pp., Budapest (Hungarian Natural History Museum).
- Norton, R. A. 1977. A review of F. Grandjean's system of leg chaetotaxy in the Oribatei (Acari) and its application to the family Damaeidae. Pp. 33–61 in: Dindal, D. L. (ed.). Biology of oribatid mites. Syracuse (SUNY College of Environmental Science and Forestry).
- & Behan-Pelletier, V. M. 2009. Suborder Oribatida. Chapter 15. Pp. 430–564 in: Krantz, G. W. & Walter, D. E. (eds). A manual of acarology. Lubbock (Texas Tech University Press).
- Okkubo, N. 2001. A revision of Oppiidae and its allies (Acarina: Oribatida) of Japan 1. Genus *Lasiobelba*. Journal of the Acarological Society of Japan 10(2): 97–109.
- Pérez-Íñigo, C. & Baggio, D. 1986. Oribates édaphiques du Brésil (III). Oribates de l'Île du "Cardoso" (deuxième partie). Acarologia 27(2): 163–179.
- Ryabinin, N. A. & Zaitsev, A. S. 2019. Two new oribatid species (Acari, Oribatida) from Sakhalin Island. Zoologichesky Zhurnal 98(4): 371–376.
- Sanjal, A. K. 1992. Oribatid Mites (Acari). Pp. 213–356 in: Ghosh, A. K. (ed.). Fauna of West Bengal. Part 3 (Arachnida and Acari). Calcutta (Zoological Survey of India).
- Sellnick, M. 1924. Einige neue südamerikanische *Damaeosoma* Arten (Acar. Oribat.). Beiträge aus der Tierkunde 1: 85–89.
- Subías, L. S. 2004. Listado sistemático, sinonímico y biogeográfico de los ácaros oribátidos (Acariformes, Oribatida) del mundo (1758–2002). Graellsia 60 (número extraordinario): 3–305.
- 2021. Listado sistemático, sinonímico y biogeográfico de los ácaros oribátidos (Acariformes: Oribatida) del mundo (excepto fósiles). 16^a actualización, 532 pp. Available from: http://bba.bioucm.es/cont/docs/RO_1.pdf
- & Balogh, P. 1989. Identification keys to the genera of Oppiidae Grandjean, 1951 (Acari: Oribatei). Acta Zoologica Hungarica 35(3–4): 355–412.
- Travé, J. & Vachon, M. 1975. François Grandjean 1882–1975 (Notice biographique et bibliographique). Acarologia 17(1): 1–19.

- Vasiliu, N. & Ivan, O. 1995. Oribatid mites from Israel. Pp. 69–86 in: Soil fauna of Israel. Bucuresti (Editura Academiei Române).
- & Ivan, O. 2011. New oppiid species (Acari, Oribatida, Oppiidae) from Romanian caves. *Travaux de l'Institut de Spéléologie "Émile Racovitza"* 50: 3–14.
- Wallwork, J. A. 1964. Some Oribatei (Acari: Cryptostigmata) from Tchad (1st. series). *Revue de Zoologie et de Botanique Africaines* 70 (3-4): 353–385.
- 1977. 4. Acarina. 4.1. Cryptostigmata. Pp. 189–257 in: La faune terrestre de L'île de Sainte-Hélène (4ème partie). *Belgique Annales, Serie 8, Sciences Zoologiques*, 220. Tervuren (Musée Royal de l'Afrique Centrale).
- Wang, H. 1993. Three new species of oppiid mites from China (Oribatida: Oppiidae). *Acta Arachnologica Sinica* 2(2): 97–103.
- Wen, Z. 1999. A new species oribatid mite of the genus *Lasiobelba* from China (Acari: Oribatida: Oppiidae). *Acta Zootaxonomica Sinica* 24(1): 46–48.